BIOMEDICAL RESEARCH SERVICE CENTER UNIVERSITY at BUFFALO, STATE UNIVERSITY of NEW YORK

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α-Ketoglutarate (α-KG) Assay Kit (Cat #: A-132)

COMPONENTS: α-KG Assay Solution- 10 ml (for 100 wells), store at -70°C

Ammonium (500x)- 0.1 ml, store at -70°C

GLDH (50x; glutamate dehydrogenase)- 0.2 ml, store at -70°C

NADPH (100x)- 0.1 ml (brown vial), store at -70°C (avoid light exposure)

20 mM α -KG- 0.2 ml, store at -20°C or -70°C

PEG Solution- 5 ml; store at 4°C (viscous; pipette solution with a cut tip)

PRODUCT DESCRIPTION: The α -KG assay is based on the difference in the UV absorption spectra between the oxidized and reduced forms of NADP+/NADPH at 340 nm (assay range $10-200~\mu$ M α -KG). The assay requires the use of UV-transparent 96-well plate or 0.1 ml quartz cuvette for measuring changes in O.D.340 nm. Reagents are stable for at least 1 year if stored and handled properly.

PROTOCOLS

Preparation of serum/plasma and cell culture medium: These samples are deproteinized by PEG precipitation. Mix 50 μ l of each sample with 50 μ l PEG Solution in a 1.5-ml microtube (PEG solution should be pipetted slowly using a cut tip). Vigorously vortex tube for at least ~30 sec to ensure thorough mixing. Keep tube on ice for 30 min. Centrifuge solution in a microfuge at ~13,000 rpm for 5 min at 4°C. Harvest supernatant and store at -20°C.

Preparation of tissue/cell samples: Cell and tissue samples are deproteinized by PEG precipitation. Please follow the extraction protocol at http://www.bmrservice.com/SupplementPEG.html. Alternatively, the samples can be deproteinized by TCA precipitation (http://www.bmrservice.com/SupplementTCA.html), which is recommended for analysis of nucleotides. Some TCA-treated samples may be diluted 3-fold with dH₂O prior to assay to achieve optimal result.

 α -KG Standards: Dilute the 20 mM α -KG standard 100-fold with dH₂O to 200 μM, e.g., 495 μl dH₂O + 5 μl 20 mM α -KG. Perform additional 1:1 dilution to generate 100 μM, 50 μM, and 25 μM α -KG standards. Store diluted standards at -20°C.

Preparation of working solution (Keep all thawed reagents on ice; do not over thaw solutions)

Each α -KG standard or sample requires 0.1 ml (or 0.2 ml; see step 4 below) working solution freshly prepared prior to assay. Calculate the amount of working solution needed for each assay. Keep working solution on ice and use immediately. An example of preparing 1 ml of working solution is given here:

 $1 \ ml \ working \ solution = 1 \ ml \ \alpha - KG \ Assay \ Solution + 2 \ \mu l \ Ammonium \ (500x) + 10 \ \mu l \ NADPH \ (100x) + 20 \ \mu l \ GLDH \ (50x)$

ASSAY:

- 1. Turn on a microplate reader or spectrophotometer. Set wavelength at 340 nm.
- 2. Prepare sufficient working solution as described above. Do not over thaw solutions during thawing, which can cause the solutions to lose sensitivity.
- 3. Pipet 20 μ l of each α -KG standard and deproteinized sample to a UV-transparent 96-well plate. Alternatively, pipette 20 μ l of each standard and sample to a set of 1.5-ml brown tubes if using a quartz cuvette.
- 4. Add 0.1 ml working solution to each well/tube followed by brief agitation or mixing. Incubate at 37°C for 60 min (do not use CO₂ incubator). For enhanced sensitivity, scale up the assay

volume by mixing 40 μl standard/sample and 200 μl working solution.

- 5. Read O.D.340 nm. Plot the α -KG standard graph as shown on the right. A new plot must be generated for each assay.
- 6. Sample α -KG concentration is derived from the trend line equation of the standard plot established at the same time (x = sample α -KG concentration in μ M; y = sample O.D._{340 nm}).

